Genomics of Ciona intestinalis

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A Functional Genomics Approach to Developmental Genetics

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Ciona intestinalis as an experimental organism

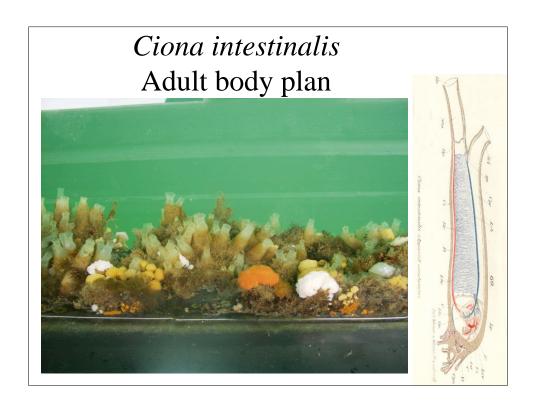


Basal Chordate
Genomic simplicity
Easy transgenics
Ectopic expression
Cis-regulation studies

Ciona intestinalis as an experimental organism

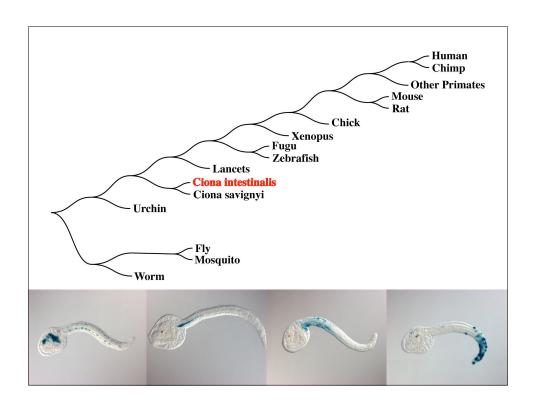


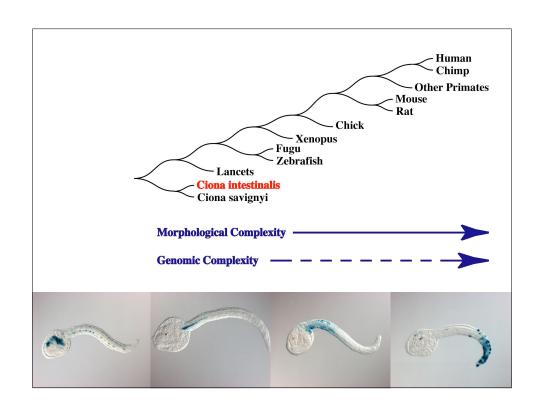
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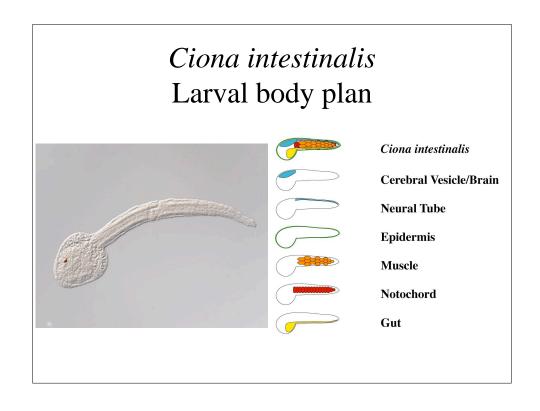




Ciona intestinalis Larval body plan Ciona intestinalis Cerebral Vesicle/Brain Neural Tube Epidermis Muscle Notochord Gut









Adult Circulatory System



• Heart

- Simple single chamber
- Muscle & pericardium
- Asymmetrical looping
- Reversible
- Vascular System
 - Two major vessels
 - Open sinuses
- Blood
 - Eight known cells types

Genomic Simplicity



- ~160,000,000bp
- ~16,000 genes
- ~1 gene every 10kb
- ~500,000 ESTs
- Small gene families
 - Predates vertebrate duplication
 - Little redundancy
 - Easily identified orthologs

Experimental Tractability



Disadvantage
No true genetics
Advantages
Easy transgenics
Scorable phenotypes
Availability

Functional Genomics

- Studying large sets of genes in parallel rather than single genes
- Experimental, not observational or modeled
- Invent new hypothesis testing experiments
- Scale traditional hypothesis testing experiments to the entire genome



Large Scale Cis-Reg Hunts

•Primary Goal

- •Screen genomic libraries for *cis*-regulatory activity
- •Catalog a large number of functionally defined *cis*-regulatory elements

Secondary Goal

•Do some targeted developmental genetics along the way



Results

Catalog a large number of functionally defined *cis*-regulatory elements

- Design, implementation and results of a small scale pilot screen of random genomic DNA 11
- Design, implementation and results of an exhaustive screen of a medium size (250kb) genomic domain
- Design, implementation of an on going large scale screen of random genomic DNA



Themes to Keep in Mind

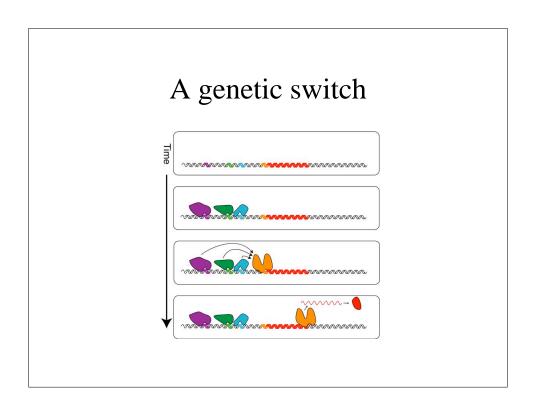
- Trade Offs
 - Number of characterized elements
 - Resolution of the characterizations
- Biases
 - Experimental biases
 - Experimenter's biases
 - Nature of enhancers vs detection methods

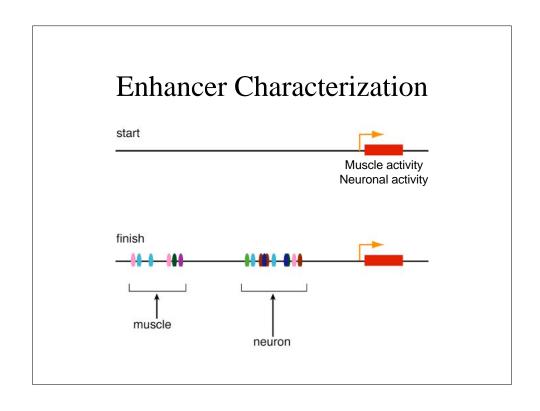


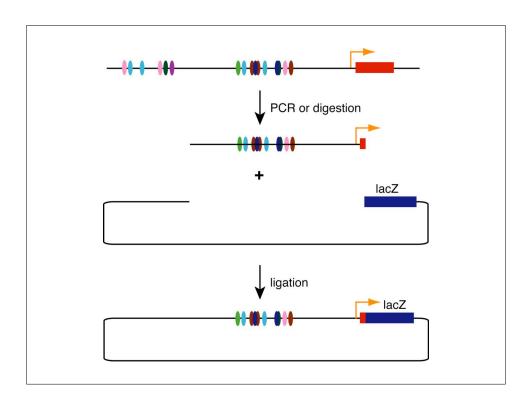
Technology to take *cis*-regulatory screening to the genomic level

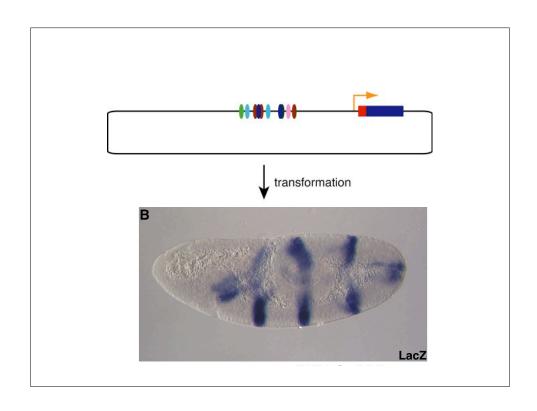
Scale traditional hypothesis testing experiments to the entire genome

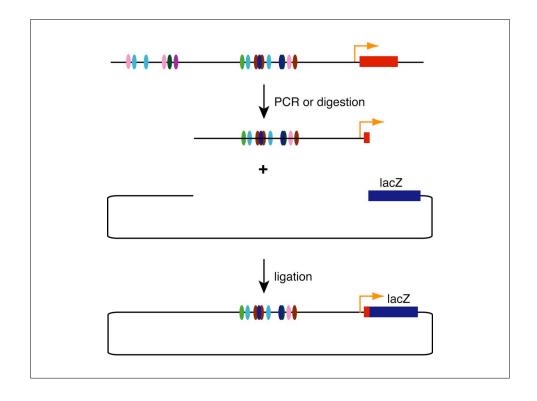


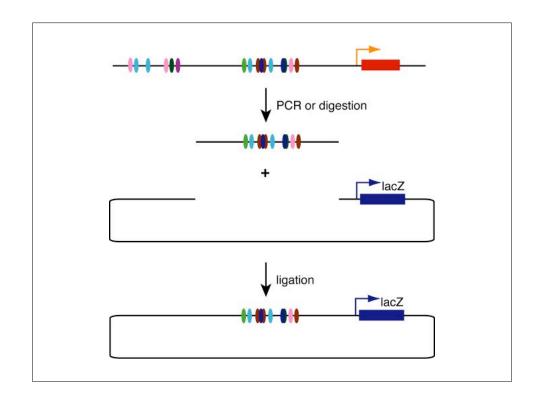


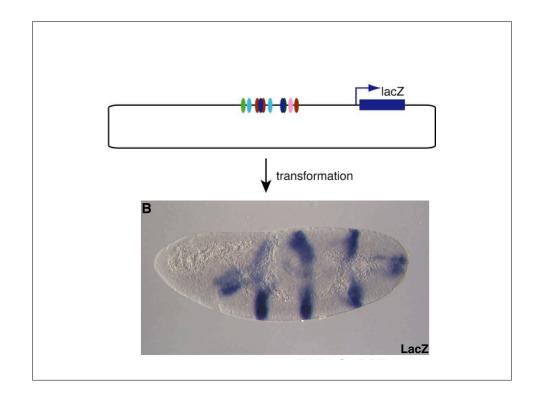












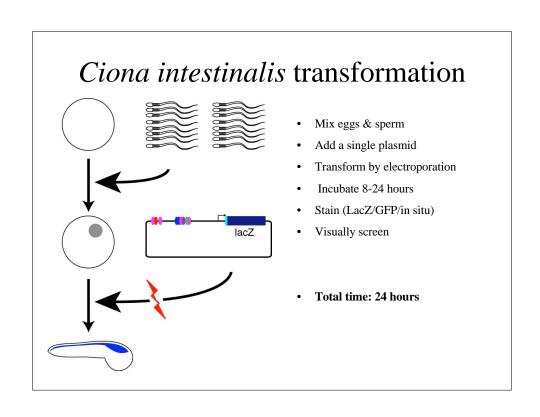
Drosophila transformation

- Collect naturally laid eggs
- Dechorionation
- Transform by single embryo microinjection
- Individually rear to 2nd generation
- Screen
- Total Time: month(s)

Traditional Enhancer Characterization

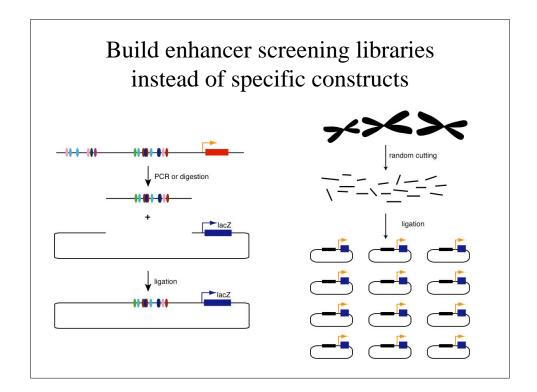
- Targeted
- Slow/Labor intensive
 - Building specific DNA constructs
 - Transforming into animals
 - Maintaining/screening animals

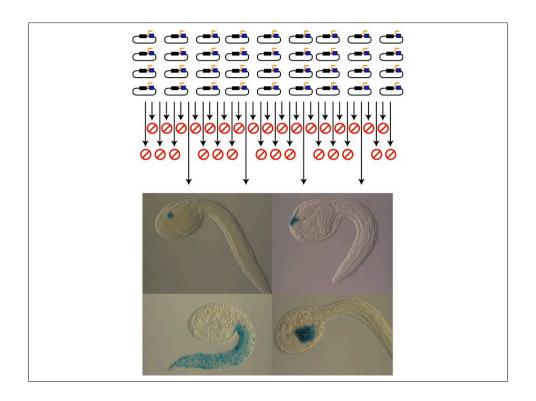




Standard *Ciona* Enhancer Characterization

- Targeted
- Slow/Labor intensive
 - Building specific DNA constructs
- Fast/Not labor intensive
 - Transforming into animals
 - Screening animals





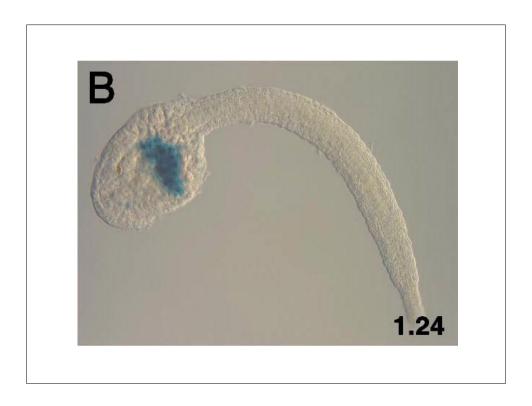
Ciona Enhancer Screening

- Non-Targeted
- Fast/Not labor intensive
 - Transforming into animals
 - Screening animals
 - Building random DNA constructs
- Limiting factors
 - DNA preps (50-100ug)
 - Transformation window (single cell embryos)
 - Imaging

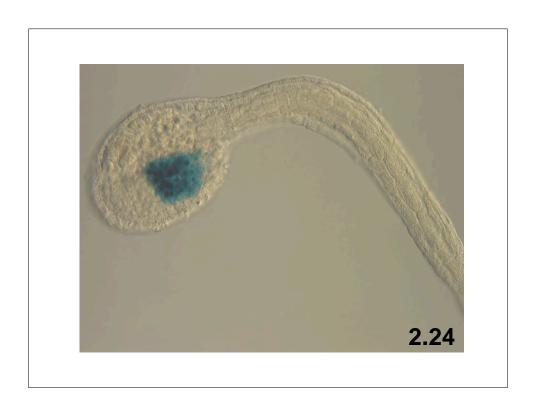
Pilot Genomic Screen

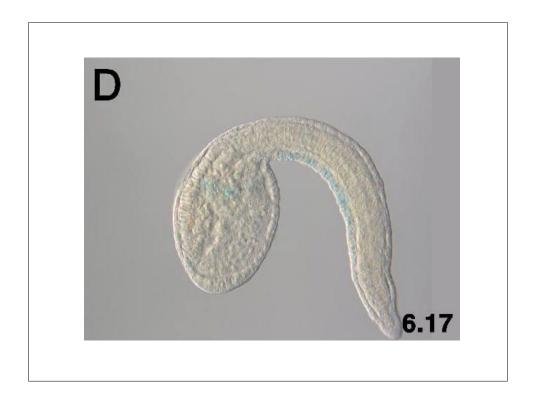
- Construct:
 - Ciona Forkhead basal promoter
 - lacZ marker detected by beta-Gal activity
 - Random genomic Sau3AI frags, 1.7kb average
- Prediction:
 - Will find cis-regulatory DNA
 - Gene density = 1 gene per 10kb. Therefore could find 1 enhancer every 10kb







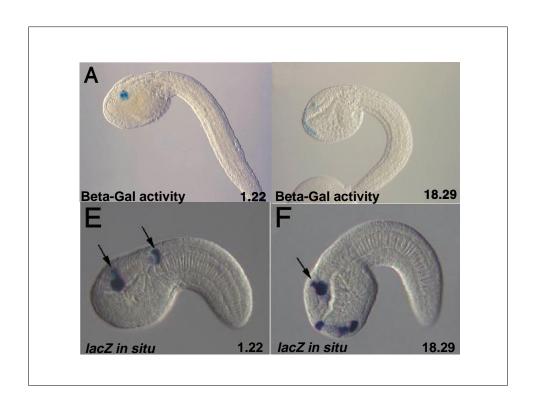




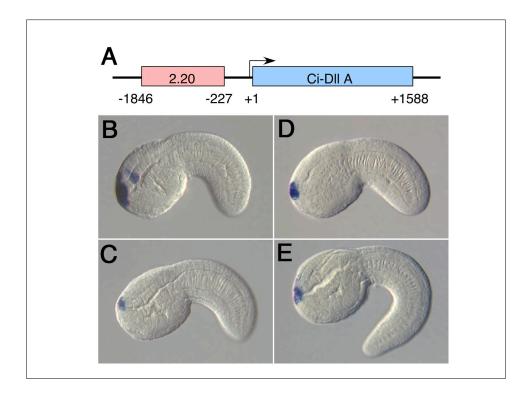


First genomic screen

- 138 constructs
- 250kb screened
- 0.15% of the genome
- Results:
- 11 strong cis-regulatory elements
- At least 8 appear to be "real" enhancer elements
- One confirmed enhancer
- 1 detectable element every 23-31 kb
- 1 detectable every 2-3 genes.







• Prediction:

- Gene density = 1 gene per 10kb
- Therefore could find 1 enhancer every 10kb

• Results:

- 1 detectable element every 23-31 kb
- 1 detectable element every 2-3 genes

Potential Issues

- Promoter specificity
- Insulators & repressors
- Enhancer Polarity
- Promoter competition
- Enhancers fragmented during cloning
- Timing
- Insufficient detection strength

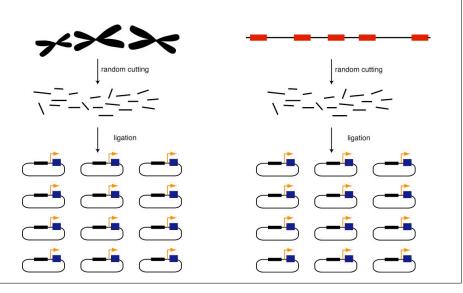
Ciona Enhancer Screening

- Non-Targeted
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 - Transforming into animals
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 - Building random DNA constructs
- Limiting factors
 - DNA preps (50-100ug plasmid)
 - Transformation window (single cell embryos)
 - Imaging

Limiting Factors

- DNA preps (50-100ug plasmid)
 - Qiagen Midipreps up to 48 constructs per day
- Transformation window (single cell embryos)
 - 24 separate constructs per batch
- Imaging
 - Quality trade offs Tough decisions

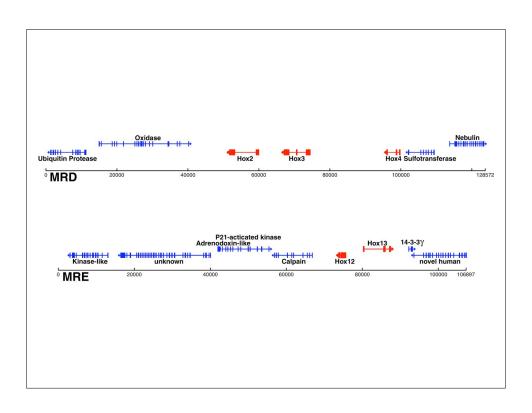
Semi-targeted *Ciona* enhancer screen Build random libraries from limited regions



Target: Ciona Hox Complex

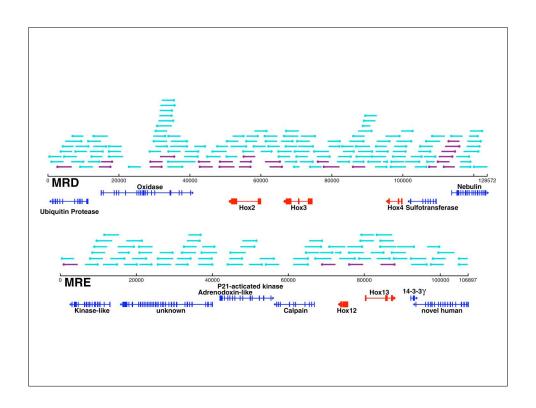
Predictions:

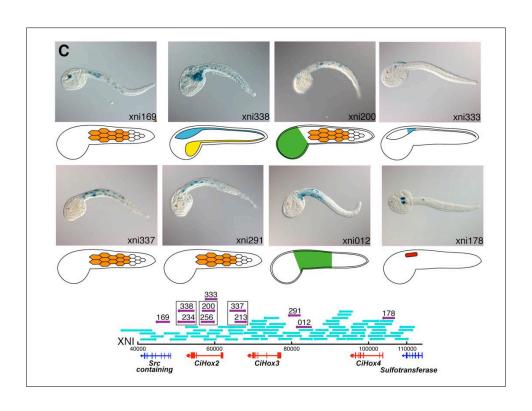
- Should be a single *Hox Complex*
- Should be a single domain
- Predictable expression patterns
 - Hox3 & Hox5 described by in situ
 (Branno & Di Lauro, Stazione Zoologica, Naples)

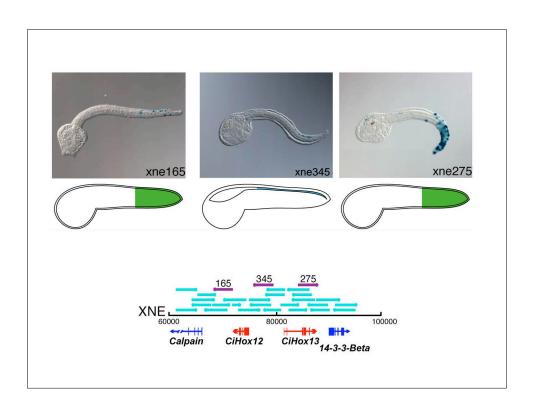


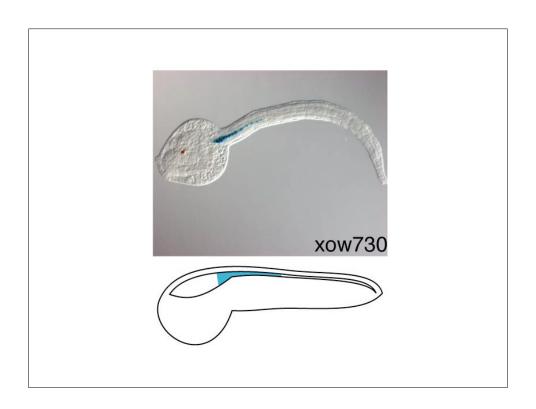
Ciona Hox Complex

- Should be a single *Hox Complex*
 - Correct
- Should be a single domain
 - Wrong, at least 4 separable domains
- Predictable expression patterns





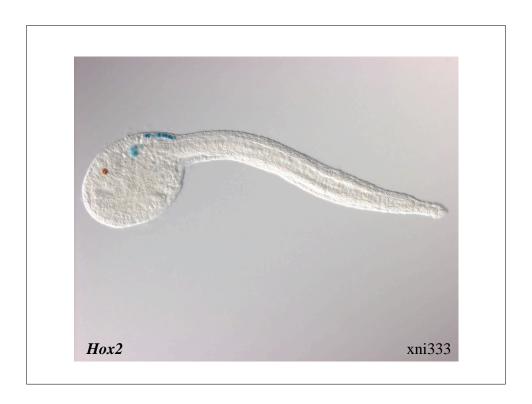


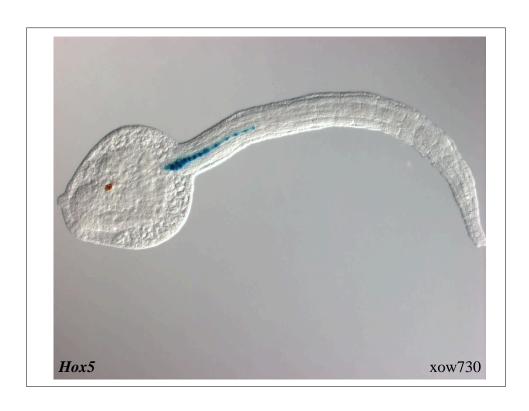


Primary results

- 221 clones electroporated & passed
- 39 clones showed positive signal
- Range for enhancers actually found:
 - Likely Maximum 30
 - Likely real21
 - Minimum 17
 - Likely *Hox* 08

















Ciona Hox Complex

- Should be a single *Hox* Complex
 - Correct
- Should be a single domain
 - Wrong, At least 4 separable domains
- Predictable expression patterns
 - Correct, Nested CNS
 - Unexpected, Nested Epidermis

Limiting Factors

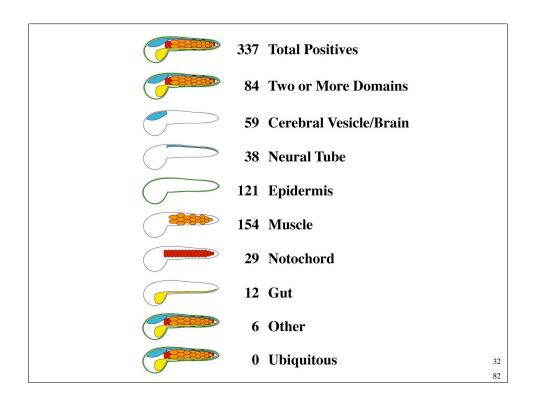
- DNA preps (50-100ug plasmid)
 - Qiagen Midipreps up to 48 plasmids per day
- Transformation window (single cell embryos)
 - 24 separate constructs per batch
- Imaging
 - Quality trade offs Tough decisions

Full Genome Scale Up



Limiting Factors

- DNA preps (50-100ug plasmid)
 - Rolling Circle Amplification
- Transformation window (single cell embryos)
 - 24 separate constructs per batch
 - 480 constructs per week
- Imaging
 - Quality trade offs Tough decisions
 - Automation??

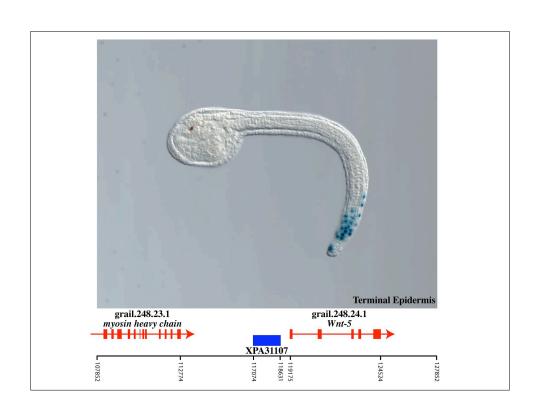


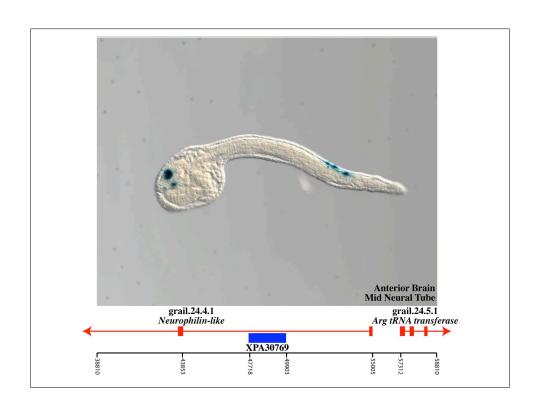
Scale Up

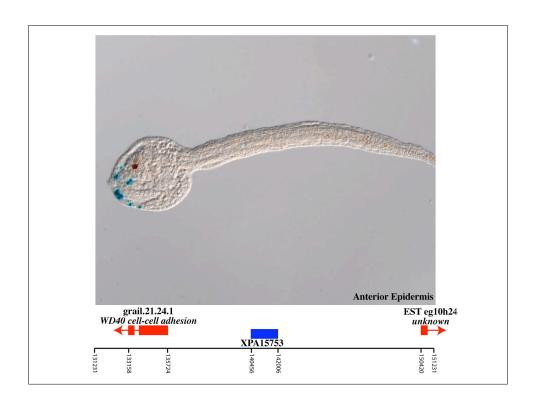
	VD 4 20106	A 11 1 1 1	Hamada i al 100 7 laba anatai a
•	XPA28186	All epidermis	Hypotheical 109.7 kDa protein
•	XPA28213	Tail Muscle	Serine/Threonine Kinase MASK
•	XPA28241	Ventral Mid Brain	Homolog to cDNA FLJ10540
•	XPA28134	Notochord	Low Sequence quality
•	XPA30404	Tail Muscle	RAR Related Steroid Receptor
•	XPA30769	Dorsal Brain, Neural Tube	e Arginine tRNA protein transferase
•	XPA30770	Muscle & Notochord	Proline Oxidase 1
•	XPA31107	Post Tail Epidermis	Wnt-2
•	XPA28831	All CSN & Epidermis	MORN motif containing
•	XPA28492	Single Cell in Brain	unknown but conserved protein
•	XPA28855	Post brain & Neural Tube	Protein kinase Ck2-beta
•	XPA29631	Neural Tube, All Gut	unknown but conserved protein
•	XPA25239	Unknown cells in head	MEC-8 like

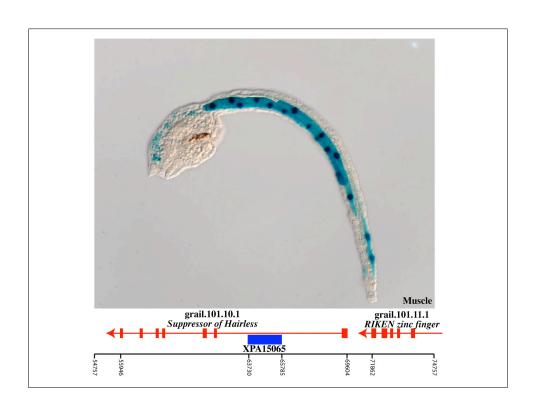
Genomic Integration

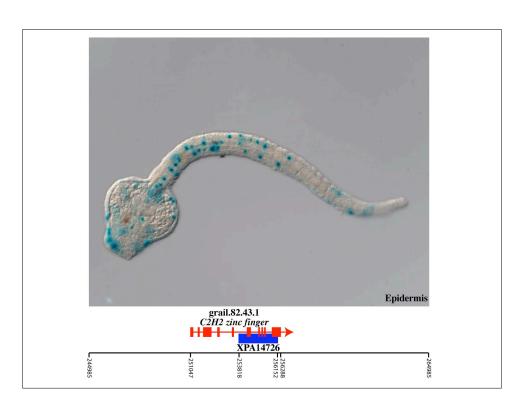
- For most random constructs, 2 end runs will
 - Identify entire subcloned sequence
 - Identify both flanking ORFs
 - Tie into EST in situ project

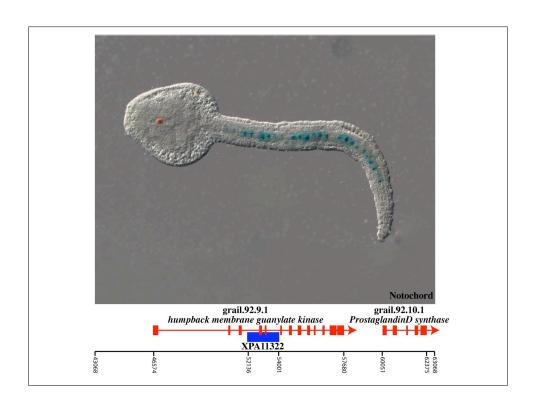


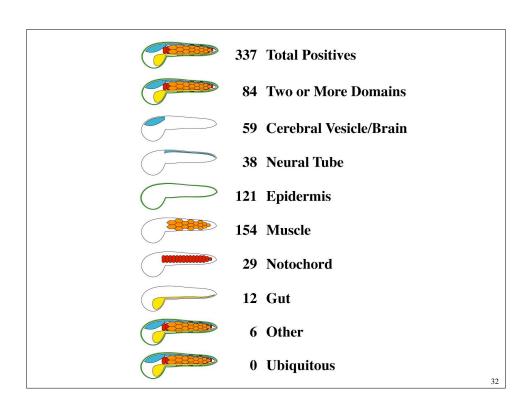


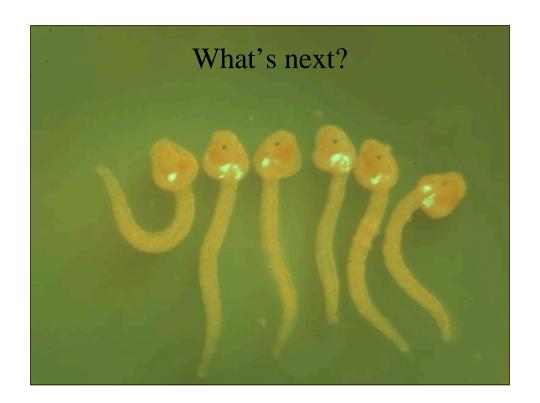












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